

THE OFFICIAL MAGAZINE OF THE OCEANOGRAPHY SOCIETY

Oceanography

CITATION

Cohen, R.E., C.C. James, A. Lee, M.M. Martinelli, W.T. Muraoka, M. Ortega, R. Sadowski, L. Starkey, A.R. Szesciorka, S.E. Timko, E.L. Weiss, and P.J.S. Franks. 2018. Marine host-pathogen dynamics: Influences of global climate change. *Oceanography* 31(2):182–193, <https://doi.org/10.5670/oceanog.2018.201>.

DOI

<https://doi.org/10.5670/oceanog.2018.201>

PERMISSIONS

Oceanography (ISSN 1042-8275) is published by The Oceanography Society, 1 Research Court, Suite 450, Rockville, MD 20850 USA. ©2018 The Oceanography Society, Inc. Permission is granted for individuals to read, download, copy, distribute, print, search, and link to the full texts of *Oceanography* articles. Figures, tables, and short quotes from the magazine may be republished in scientific books and journals, on websites, and in PhD dissertations at no charge, but the materials must be cited appropriately (e.g., authors, *Oceanography*, volume number, issue number, page number[s], figure number[s], and DOI for the article).

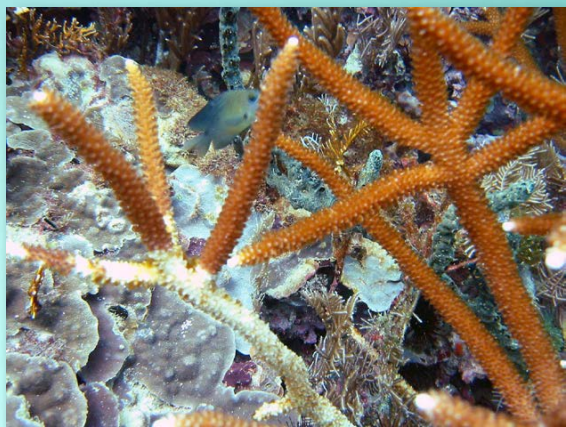
Republication, systemic reproduction, or collective redistribution of any material in *Oceanography* is permitted only with the approval of The Oceanography Society. Please contact Jennifer Ramarui at info@tos.org.

Permission is granted to authors to post their final pdfs, provided by *Oceanography*, on their personal or institutional websites, to deposit those files in their institutional archives, and to share the pdfs on open-access research sharing sites such as ResearchGate and Academia.edu.

MARINE HOST-PATHOGEN DYNAMICS

Influences of Global Climate Change

By Rebecca E. Cohen, Chase C. James, Allison Lee, Maurizio M. Martinelli, Wendy T. Muraoka, Mikayla Ortega, Rachel Sadowski, Lark Starkey, Angela R. Szesciorka, Sonya E. Timko, Elliot L. Weiss, and Peter J.S. Franks



ABSTRACT. Host-pathogen relationships are an important subset of interspecies interactions that are capable of influencing population dynamics, community structure, and biogeochemical cycles. Relationships between hosts and pathogens are expected to shift in response to global climate change. Atmospheric and oceanic warming is already affecting many ecosystems, perhaps most markedly those in marine environments. Case studies illustrate some impacts ocean warming is having on marine host-pathogen dynamics, including: (1) increased host stress, (2) increased pathogen virulence, (3) pathogen range expansion, and (4) host range changes. Output from global climate models was used to calculate the number of additional months each year that various coastal regions around the globe may reach temperatures above a given threshold 50 and 100 years from now; for the Arctic region, we characterize changes in sea ice cover. Complex interactions between multiple ocean conditions that could also affect host-pathogen relationships, such as rising sea surface temperature, anoxia, ocean acidification, and stratification, should also be investigated.

INTRODUCTION

Current scientific consensus identifies anthropogenic carbon emissions as the major cause of the unprecedented rates of atmospheric and oceanic warming observed over the last few decades (IPCC, 2014). Between 1971 and 2010, the upper 75 m of the ocean warmed by 0.11°C per decade, while snow and ice cover concurrently diminished (IPCC, 2014). Such trends are predicted to continue throughout the rest of this century. By the year 2100, sea surface temperatures are expected to warm by 1.1°–2.9°C based on the most conservative B1 emission scenario, and by 2.0°–5.4°C based on the more realistic A2 scenario (IPCC, 2007), with warming continuing beyond 2100 under all predicted scenarios (IPCC, 2014). This warming is considered irreversible on a multicentury timescale unless efforts are made to actively remove CO₂ from the atmosphere (IPCC, 2014).

The impacts of these changes are already apparent in many ecosystems, but perhaps most markedly in marine environments, particularly through increased durations of high-temperature events. The 2016 mass bleaching of corals on the Australian Great Barrier Reef, for example, resulted from the cumulative impacts of several anomalously warm months (Hughes et al., 2017). Such cumulative effects are expected to manifest

themselves as changes in many aspects of marine ecosystems, including total biomass, community structure, diversity, and species interactions—as well as changing host-pathogen dynamics. Here, we explore the cumulative impacts of temperature changes on host-pathogen dynamics, using examples from the poles to the equator.

The persistent trends in environmental variables driven by global climate change motivate scientists to quantify the effects of environmental and physiological changes on host organisms and pathogens (e.g., Maynard et al., 2016). Changes in the environment can result in outcomes that favor either host or pathogen, increasing or decreasing local infection rates (Danovaro et al., 2011; Burge et al., 2014). Hosts may exhibit impaired immune response, habitat loss, or range expansion, while pathogens may develop increased virulence, pathogenicity, expanded ranges, or decreased range overlap with their hosts (Harvell et al., 2002). Better understanding of these dynamics will improve our ability to predict changes in host-pathogen interactions and the expected impacts on ecosystem structure and composition. At an ecosystem scale, these impacts can cascade through trophic pathways, affecting organisms from primary producers to top predators.

Changes in temperature, pH, O₂, sea ice extent, and many other environmental variables contribute to the impact of climate change on disease in marine systems. Here, we use temperature and sea ice cover as the primary variables through which we explore changes in host-pathogen relationships. Temperature accounts for much of the spatial and temporal biogeographic structure of the ocean (Kordas et al., 2011) through its effects on the growth, survival, health, and reproduction of marine organisms (Doney et al., 2012). Our decision to use temperature as an indicator variable was also governed by the availability of measured temperature responses for many taxa (both hosts and pathogens alike) and global model temperature projections for the next hundred years.

Temperature is also a particularly useful proxy for global climate change. Furthermore, many hosts display cumulative responses to temperature trends. For example, cumulative heat stress—rather than any single daily temperature—is a primary driver of bleaching for many corals (Berkelmans 2002; Heron et al., 2016; Hughes et al., 2017). We use output from global climate models to explore the effects of climate change on selected host-pathogen systems. We quantify changes in the annual cumulative duration of sea surface temperatures and sea ice concentrations relative to thresholds relevant to particular host-pathogen interactions. Using case studies as examples, we explore four fundamental modes through which climate change is likely to affect host-pathogen dynamics.

1. Increased Host Stress. Many marine taxa exhibit stress in response to changes in their physical environments (Burge et al., 2014). Small increases in global ocean temperature could lead to broad-scale changes in stress-induced host immune responses (Harvell et al., 2002). With their global distributions, sea

urchins and their pathogens serve as useful models for this response (Maes and Jangoux, 1984; Lester et al., 2007; Becker et al., 2008).

2. Increased Pathogen Virulence. Elevated temperatures increase the virulence of many marine pathogens (Burge et al., 2014) by increasing pathogen metabolism and fitness, ultimately leading to higher rates of transmission (Karvonen et al., 2010). The pathogen *Vibrio coralliilyticus* and one of its coral hosts, *Pocillopora damicornis*, provide a useful case study of increased virulence in the face of warming ocean temperatures driven by anthropogenic climate change. *Vibrio* species are globally distributed and can infect a wide range of organisms. The increasing success of *Vibrio* in a changing ocean may have important implications for marine community composition (Ben-Haim et al., 2003a; Kimes et al., 2012; Vezzulli et al., 2013).

3. Pathogen Range Expansion. Regionally, warming seas may allow pathogens to expand farther into host habitat (de Montaudouin et al., 2010; Xu et al., 2017). The well-studied interaction between the parasite *Perkinsus marinus* and its host, the oyster *Crassostrea virginica*, is a useful example of this dynamic. This system is representative of host species whose ranges currently extend beyond the ranges of their

pathogens; with increasing temperatures, pathogen ranges are likely to encroach into those of unaffected populations.

4. Host Range Changes. Shifts in species distributions as a result of altered habitat may increase exposure rates and transmission of certain pathogens, or introduce novel pathogens into a system. This can be seen in the interactions between *Brucella* spp. and their marine mammal hosts, such as pinnipeds. Temperatures in polar regions, where most pinniped species are found, are increasing at nearly twice the global average rate, resulting in substantial reductions in ice cover (IPCC, 2007; Kovacs, 2011). These changes will manifest in different outcomes for species that depend upon sea ice versus species that avoid it. Range restriction of ice-dependent species has the potential to increase local population densities, which is likely to favor increased transmission of pathogens within populations. Simultaneously, range expansion of species that avoid polar ice may facilitate range expansion of their pathogens, leading to increased transmission among populations.

METHODS

Climate projections used here were obtained from A2 scenario data made available by the Intergovernmental Panel on Climate Change (IPCC) Fourth Assessment SRES AR4 CMIP3

experiments (Meehl et al., 2007). The data are freely available from the IPCC Data Distribution Center (<http://www.ipcc-data.org>). The A2 scenario emphasizes economic and global integration over environmental or regional solutions. It is characterized by a more divided world (i.e., more independently operating, self-reliant nations), continuously increasing population, and slower economic and technological development compared to other scenarios (IPCC, 2007). The A2 scenario projects a best-estimate global surface temperature rise of 3.4°C with a likely range of 2.0°C to 5.4°C by 2100. Models for sea surface temperature and sea ice coverage were selected based on their availability and ease of online access. Monthly sea surface temperature projections were generated by the ECHAM5/MPI-OM model from the Max-Planck Institute for Meteorology (Hamburg, Germany). The model had a spatial resolution of 1.9° × 1.9° and is described in detail by Roeckner et al. (2004). The ECHAM5/MPI-OM model from 2000 to 2100 projects a global sea surface temperature rise of 2.5°C (Figure 1).

Using the ECHAM5/MPI-OM model, average sea surface temperatures from 2001 to 2017 were calculated to use as a baseline (Figure 2a–c) against projected sea surface temperatures in 2050 and 2100 (Figure 2d–i), averaged over a five-year span to account for natural annual variability. From these data, we calculated the number of months each year during which temperature was greater than a given threshold: 24°C, 27°C, or 30°C. We then subtracted the baseline (2001–2017) from the projected number of months exceeding the temperature thresholds. These yearly anomalies were used to make predictions concerning the effects of global climate change on various aspects of host-pathogen relationships in different regions. All plots were made in MATLAB (version 2016b) using `m_map` (version 1.4b).

Monthly sea ice coverage projections were obtained from the CM2.0 model

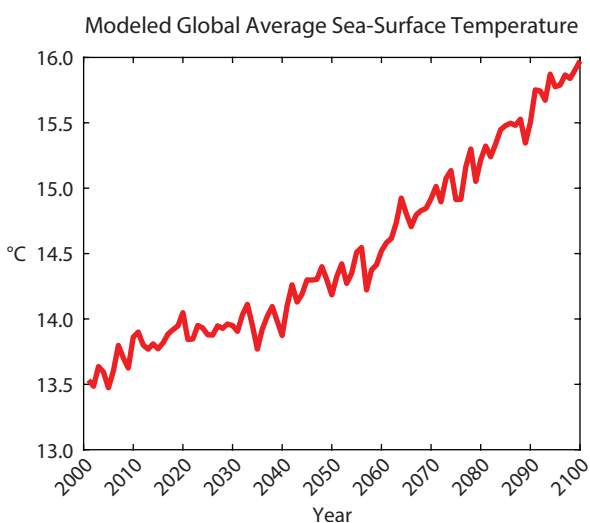


FIGURE 1. Projected sea surface temperatures from 2000 to 2100 from the ECHAM5/MPI-OM model using the A2 climate scenario (Meehl et al., 2007).

of the Geophysical Fluid Dynamics Laboratory (GFDL; Princeton, NJ, USA). The GFDL CM2.0 model has an ocean resolution of $0.3^{\circ}\text{--}1.0^{\circ} \times 1.0^{\circ}$ with depth as the z -coordinate and an explicit free surface top boundary condition. Gnanadesikan et al. (2006) provide a detailed description of the model, including benefits and drawbacks of the model parameters. Ice area concentrations of 75% and 90% are used to compare seasonal changes in ice area for 2017, 2050, and 2100, and mapped according to periods of known haul-outs for ringed (March–May) and fur (June–July) seals.

CASE STUDIES

Increased Stress Inhibits Host Recovery

Urchins, present in all ocean basins and occupying a variety of temperature ranges, may represent an indicator for increases in host infections due to thermal stress-induced declines in immune responses. Bald sea urchin disease is an umbrella term describing an assortment of urchin diseases that trigger similar symptoms and that are caused by a variety of pathogenic bacteria (Jangoux, 1987; Lester et al., 2007; Becker et al., 2008). The bacteria consume healthy urchin tissue to form distinct lesions that range in size from a few

millimeters to more than one-third of the host's surface, with many urchins afflicted by multiple lesions (Maes and Jangoux, 1984; Jangoux, 1990; reviewed by Becker et al., 2008). Regeneration and recovery from bald sea urchin disease is possible; however, in cases where the skeleton is breached or a majority of the urchin is covered in lesions, death is common (Maes and Jangoux, 1984; Jangoux, 1987, 1990; reviewed by Becker et al., 2008).

Infection rates for bald sea urchin disease in both tank experiments and multiple real-world mass mortality events are strongly correlated with increases in temperature (Scheibling and Stephenson, 1984; Ward and Lafferty, 2004; Girard et al., 2012; Brothers et al., 2016; Sweet et al., 2016). At higher temperatures, urchins experience thermal stress, which initially triggers a heightened immune response (Brothers et al., 2016). Despite these immunological responses, increased infection rates often occur when elevated temperatures persist or are more extreme. Furthermore, urchins often exhibit decreased activity and feeding during extreme heat events, as well as decreased fecundity, which can restrict recovery of individuals and populations (Farmanfarman and Giese, 1963; Lester et al., 2007). While negatively impacting the urchins, these extreme heat events

appear to favor the pathogens, increasing growth and transmission rates, as well as improving bacterial survivorship (Brothers et al., 2016; Sweet et al., 2016). The longer the urchins remain in a state of stress, the higher their susceptibility to disease (Brothers et al., 2016; Sweet et al., 2016). Though recovery is possible, urchin species impacted by a bald sea urchin disease event tend to exhibit increased mortality in persistently elevated seawater temperatures (Lester et al., 2007; Girard et al., 2012; Brothers et al., 2016).

Critical temperatures above which host immune responses decrease are 23°C for *Strongylocentrotus purpuratus*, *Helicidaris erythrogramma*, and *Strongylocentrotus intermedius* (Farmanfarman and Giese, 1963; Agatsuma, 2007; Brothers et al., 2016) and 29°C for *Paracentrotus lividus* (Boudouresque and Verlaque, 2007). We predict that the number of months exceeding each species' critical temperature will increase from the baseline through 2100 (Figure 3). These increases will grow spatially and become longer in duration over the next 100 years in both temperate and subtropical regions.

Predicted increases in the frequency, duration, and intensity of warm-water events suggest reduced survival and recovery from future mass mortality events for many urchin populations.

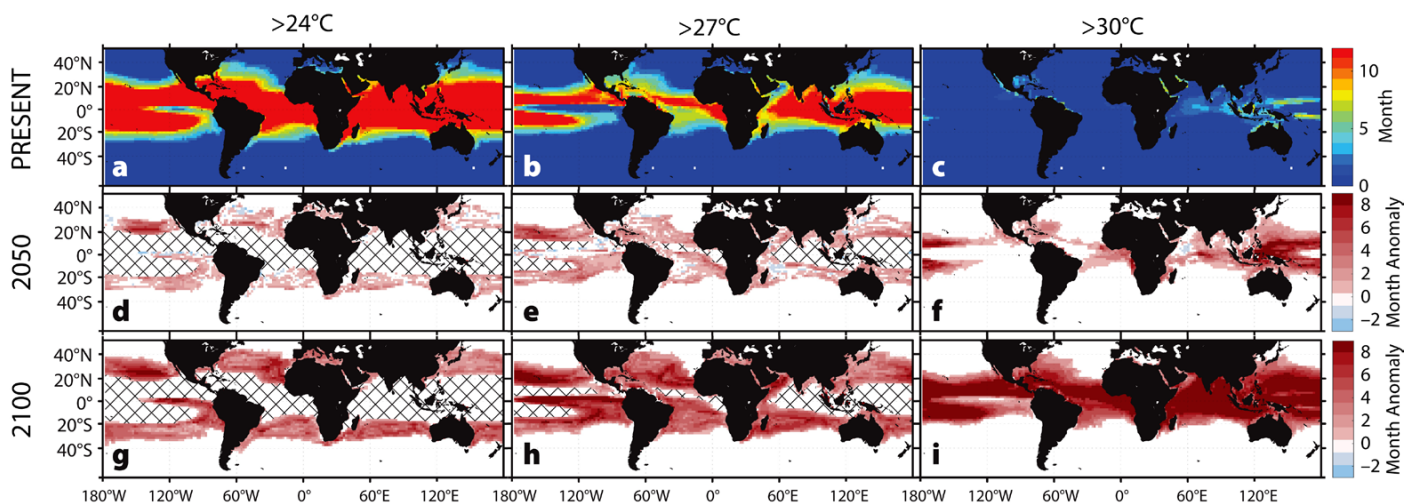


FIGURE 2. Predicted increases in the duration of warm-water events. (a–c) Average number of months sea surface temperatures exceeded the given temperature threshold (Present: 2001–2017). The baseline durations (a–c) were subtracted from projected durations in 2050 (d–f) and 2100 (g–i) to give predicted annual anomalies in the duration of warm-water events. Cross-hatching denotes areas where all 12 months exceeded the threshold temperature duration of the baseline.

For example, two to four more months of above-critical temperatures each year in the Mediterranean Sea will affect *P. lividus* by 2100 (Figure 3d,h,l). In coastal regions, up to five additional months of temperatures above 30°C are predicted, a trend that may lead to local extinctions. Similarly, *S. intermedius* may be exposed annually to two more months of above-critical temperatures around Japan and the Korean peninsula by 2100 (Figure 3a,e,i). In the global warming hotspot off the east coast of Australia, home to *H. erythrogramma*, at least four to six more months of above-critical temperatures are predicted by 2100, with some regions expected to increase by seven or eight more months (Figure 3b,f,j). The most marked temperature increases occur in the range of *S. purpuratus* along the west coast of North America, with seven to eight more months of elevated temperatures predicted for some areas each year (Figure 3c,g,k). The region extending across the Pacific between 16°N and 36°N may exceed 27°C for eight additional months per year, with potentially

extreme consequences for resident populations (Figure 2).

These predicted increases in the frequency, duration, and intensity of warming events will cause urchins—and many other organisms living within these areas—to respond to enhanced thermal stress with altered immune responses and increased mortality rates due to pathogenic infection.

Increased Temperature Increases Pathogen Virulence

Vibrio are among the most common pathogens in the marine environment (Vezzulli et al., 2013) and comprise the largest genus (81 species) in the Vibrionaceae family (Thompson et al., 2007). In corals alone, geographically distinct strains of *V. coralliilyticus* can cause specific infections and diseases based on host, location, and local temperature (Pollock et al., 2010).

Pocillopora damicornis is found throughout the Pacific and Indian Oceans and in some areas serves as the dominant reef engineer (Combosch and Vollmer,

2011; Hoeksema et al., 2014). Most scleractinian corals rely for a significant portion of their energy requirements on intracellular, symbiotic, photosynthetic dinoflagellates (“zooxanthellae” primarily of the genus *Symbiodinium*; Bourne and Munn, 2005; Bourne et al., 2009). *V. coralliilyticus* is known to infect *P. damicornis*, impacting both the coral host and its zooxanthellae (Ben-Haim et al., 2003b).

The host-pathogen interactions of *P. damicornis* and *V. coralliilyticus* are critically dependent on seawater temperature. At 25°C, *V. coralliilyticus* becomes infectious to *P. damicornis* (Ben-Haim et al., 2003b). Above 27°C, pathogen virulence increases significantly, coral fitness is reduced, and *P. damicornis* is no longer able to adequately resist infection. In addition, multiple virulence factors, including transcriptional regulation, quorum sensing, antimicrobial resistance, and toxin secretion systems, are upregulated at this temperature (Kimes et al., 2012). Extracellular proteases produced by *V. coralliilyticus* can cause subsequent lysis and mortality of

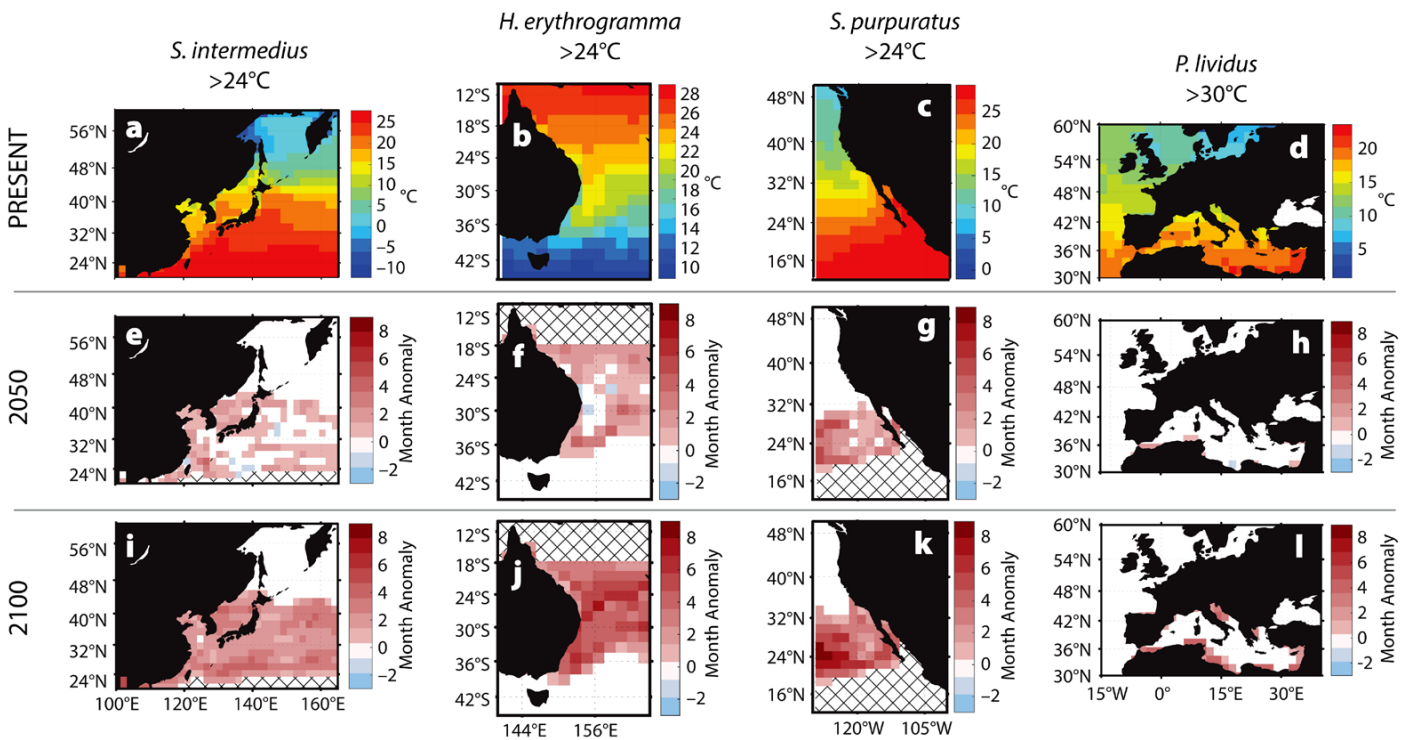


FIGURE 3. Predicted increases in the duration of warm-water events for four species of urchin from four geographic areas affected by bald sea urchin disease. (a–d) Baseline temperature conditions (2001–2017). (e–h) Changes in the number of months the temperature is predicted to exceed the given threshold by 2050, and (i–l) by 2100. Cross-hatching same as Figure 2.

P. damicornis (Ben-Haim et al., 2003a), which, in laboratory studies, occurs within two weeks at temperatures above 27°C (Ben-Haim et al., 2003b).

The tropics (23.5°S to 23.5°N) already exhibit consistent annual temperatures of 24°C to 27°C (Figure 2a,b), and that range will expand into the future. Heat-stressed corals and their symbionts release higher concentrations of dimethylsulfoniopropionate (DMSP), which acts as a trigger for chemotaxis (directional movement in response to a chemical stimulus) and chemokinesis (a change in swimming behavior in response to a change in chemical concentration) in *V. coralliilyticus* (Garren et al., 2014, 2016). Upon entry to the host cells, *V. coralliilyticus* targets the *P. damicornis* zooxanthellae (Ben-Haim et al., 2003a), resulting in the expulsion of the symbionts and consequent bleaching (Dove and Hoegh-Guldberg, 2006). Recovery may be possible if temperatures return to an acceptable level for a long enough period. However, with projected increases in sea surface temperatures for 2050 (Figure 2d–f) and 2100 (Figure 2g–i), mass coral mortalities can be expected (D’Croz and Maté, 2004).

With elevated sea surface temperatures, we can expect to observe shifts in the dynamics of *V. coralliilyticus* and *P. damicornis* (Ben-Haim et al., 2003a,b). At temperatures greater than 30°C, *V. coralliilyticus* abundance increases by up to four orders of magnitude relative to 24°C, further increasing the likelihood of infection (Tout et al., 2015). Climate models predict that by 2050, a portion of the Indo-Pacific will experience up to seven additional months of temperatures greater than 30°C (Figure 4b), and by 2100 at least eight additional months of temperatures greater than 30°C annually for the entire region (Figure 4c).

Most of the tropics are already near or above these threshold temperatures for the majority of the year (Figure 2), and these conditions are expected to expand poleward. As a result, the spatial extent of certain highly virulent pathogens may expand as climate change progresses.

Temperature Facilitates Pathogen Range Expansion

While the virulence of local pathogen populations is likely to increase, we also expect to see pathogen range expansion in response to increasing temperatures. To explore this possibility, we considered the single-celled protozoan *Perkinsus marinus* and its economically important eastern oyster host *Crassostrea virginica*. Oysters are often used as indicators of environmental health—their efficient filter-feeding mechanisms increase the exposure of their tissues to pathogens in the surrounding waters (Segarra et al., 2010; Jenkins et al., 2013; Bidegain et al., 2016). Oysters are also globally distributed, with a wide range of temperature tolerances (Soudant et al., 2013); the spread of *P. marinus* to *C. virginica* has historically been restricted by the temperature threshold of the pathogen.

Originally classified as the fungus

Dermocystidium marinum, which led to the disease name “dermo” (Bower et al., 1994), *P. marinus* infects several mollusc species but has been most extensively studied in *C. virginica* (Bower, 2013). *P. marinus* competes with its oyster host for nutrients and is known to cause disease and mortality (Bower et al., 1994; Soudant et al., 2013). The infected oyster becomes stressed, tissues pale in color, and gamete production and growth slow. Emaciated, the mantle shrivels and pulls away from the shell, and pockets of pus-like fluid develop (Bower et al., 1994). Lysis of tissues and blockage of blood vessels cause fatality, though many oysters can persist up to three years with active infections (Sunila, 2007). Adult oysters can tolerate sustained temperatures of 2°C to 36°C, and up to 49°C for short intervals (Banks et al., 2007), with the optimal range between 20°C and 30°C. This wide temperature tolerance allows oysters to

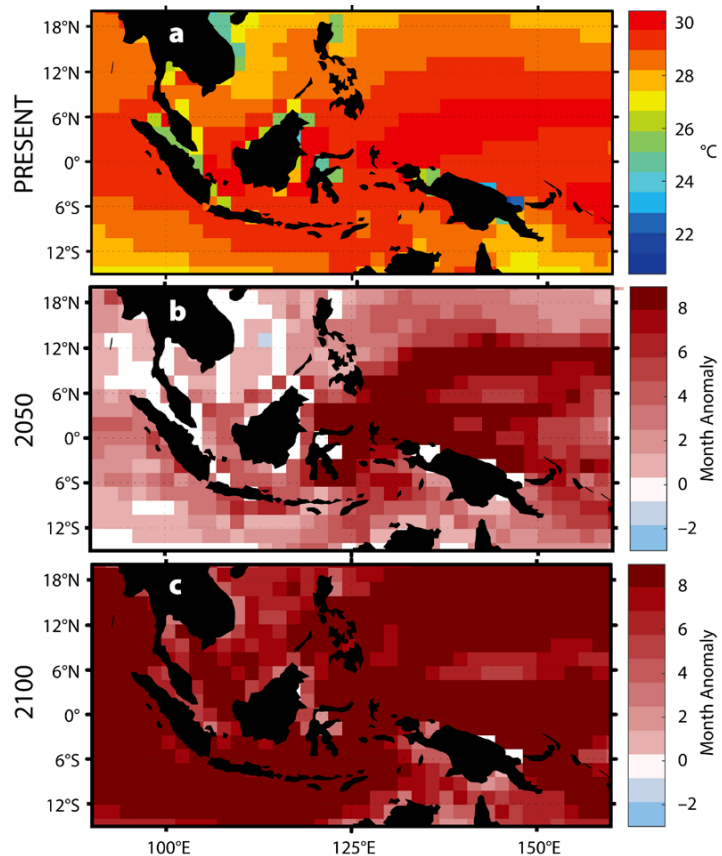


FIGURE 4. Predicted changes in the thermal habitat of *V. coralliilyticus*. (a) Present annual average temperatures. (b–c) Predicted changes in the duration of warm-water events (>30°C) in (b) 2050 and (c) 2100.

inhabit the majority of the world's coastlines (Banks et al., 2007). The temperature tolerance of *P. marinus* overlaps that of *C. virginica*, with growth between 18°C and 20°C, increase in infection intensity above 20°C, and highest infection intensity as temperatures near 25°C (La Peyre et al., 2008). *P. marinus* can survive down to 4°C (La Peyre et al., 2008).

Along the east coast of the United States, our analyses predict longer periods

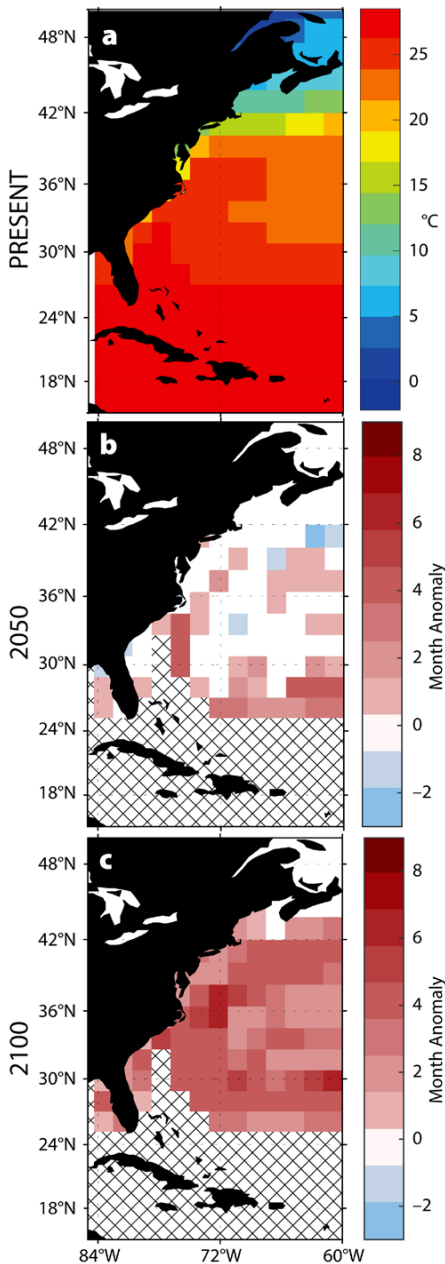


FIGURE 5. Predicted changes in the thermal habitat of *P. marinus*. (a) Present annual average temperatures. (b–c) Predicted changes in the duration of warm-water events (>24°C) in (b) 2050 and (c) 2100.

of sustained elevated temperatures, which may allow the range of *P. marinus* to expand into the established habitat of *C. virginica* and increase the potential for infection (Figure 5). Annual durations of ocean temperatures greater than 24°C are predicted to increase up to four months by 2050, and up to six months by 2100, in this region. The long duration of elevated temperatures creates the potential for pathogens to become established farther north along the coast. This potential for pathogen range expansion is corroborated by changes during the 1990–1992 warming period when sea surface temperatures increased by 0.27°C to 0.77°C per year along the US east coast (Cook et al., 1998). The subsequent *P. marinus* expansion devastated *C. virginica* populations, causing damage to previously unaffected oyster populations. Some populations suffered infection rates of 90%–100% of individuals, with overall mortality reaching 60%–80% (Ford and Smolowitz, 2007).

Range expansions of pathogens into new areas where hosts may be unaccustomed to infection can thus result in unusually high mortality events. As these range shifts transition from episodic to permanent, relationships between hosts and their pathogens are likely to change.

Biogeographic Shifts Increase Incidence of Infection

Due to the disproportionately higher rate of temperature change observed at the poles, organisms such as pinnipeds may exhibit substantial biogeographic shifts in response to changes in suitable habitat availability. Pinnipeds are widely distributed throughout Arctic and Antarctic waters, with some species extending far into the subtropics (Pompa et al., 2011). Because pinnipeds must haul out on land or ice for breeding and pupping, they are at risk of exposure to both marine and terrestrial pathogens. *Brucella*, the causative agent of brucellosis, is a genus of gram-negative coccobacillus bacteria found in both marine and terrestrial environments. *Brucella* grows between

20°C and 40°C and optimally at 34°C (de Figueiredo et al., 2015). *Brucella* species do not multiply outside their hosts, but can persist for months in 10°C to 15°C conditions and for years in frozen tissue (Nymo et al., 2011). Scholz et al. (2008) demonstrated that one species, *B. microti*, has also found a long-term reservoir in soil. Because this wide temperature range enables *Brucella* to survive throughout the world ocean, instead of considering the direct impacts of Arctic temperature change on *Brucella* survival, we focused on changes in intra- and inter-population transmission rates as a result of sea ice loss.

Ten species of *Brucella* are recognized, with *B. pinnipedialis* and *B. ceti* primarily affecting their eponymous marine mammal groups. High *Brucella* seroprevalence rates have been observed among several pinniped species, such as hooded seals (*Cystophora cristata*) in the North Atlantic and Barents Sea (35%–38%), and common seals (*Phoca vitulina*) along the coast of Scotland (49%; Spickler, 2009). Brucellosis can present a wide variety of clinical symptoms: infection of reproductive tissue resulting in placentitis and spontaneous abortion; neurobrucellosis, associated with tremors, seizures, and strandings; and damage to tissue and joints as well as respiratory problems caused by systemic brucellosis (Thakur, 2012). *Brucella* spp. have also been found in the tissues of apparently healthy individuals (Spickler, 2009).

There have been multiple reported cases of *Brucella* infecting species other than their preferred hosts, and laboratory studies suggest that *B. pinnipedialis* has the potential to infect a wider range of animals (Lambourn et al., 2013). Vertical transmission is thought to occur during gestation and lactation, while horizontal transmission may take place via contact with infected individuals or their aborted fetuses, placentas, birth fluids, and secretions (Thakur et al., 2012).

Diminishing sea ice cover in the Arctic may increase occurrences of brucellosis among pinnipeds by increasing local

■ Northern Fur Seal Rookeries (Ice-Avoiding)

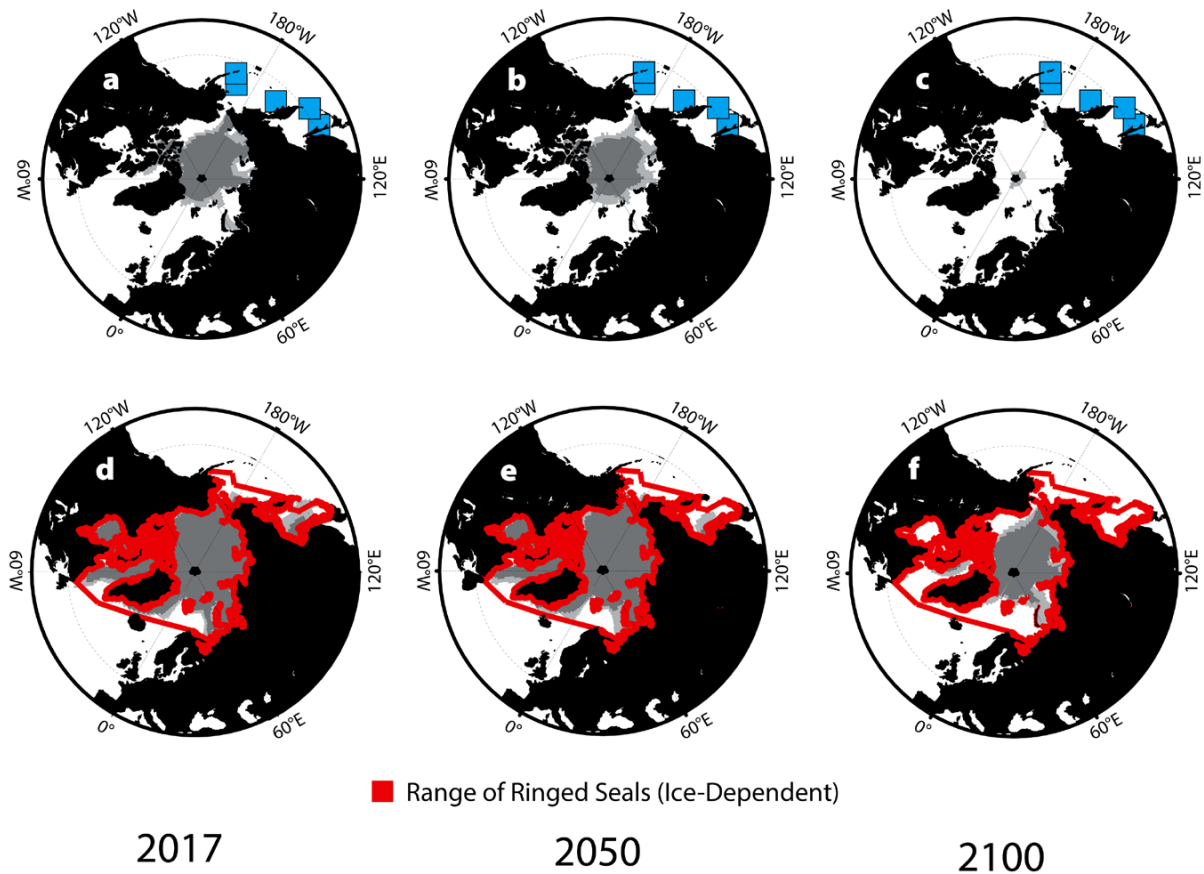


FIGURE 6. Habitat ranges for pagophilic and land-breeding species throughout the North Pacific with 75% and 90% ice cover (light and dark gray, respectively) for 2017, 2050, and 2100. (a–c) Island rookeries for the northern fur seal (*Callorhinus ursinus*) from June–July. (d–f) Peak habitat range for ringed seals (*Phoca hispida*) from March to May.

population densities of pagophilic (ice-dependent) seals. Most of these species currently haul out on ice in low densities, generally fewer than one to two animals per square kilometer (Young et al., 2015). Using predicted Arctic ice cover during the pupping season from the GFDL CM2.0 model, we looked at changes in reproductive habitat for ice-dependent species (Figure 6). Based on our predictions, substantial reductions in ice and snow available for excavating subnivean birth dens are likely to increase competition for prime whelping, breeding, and molting sites among pagophilic species. Greater densities and increased contact among reproductive females may increase transmission of *Brucella*. For species that almost never haul out on land and prefer habitat with 75%–90% ice coverage (e.g., bearded seals, *Erignathus barbatus*,

and ringed seals, *Phoca hispida*), this habitat constriction is particularly apparent (Figure 6; Simpkins et al., 2003).

Brucellosis may also spread following range expansion of land-reproducing pinnipeds that exploit exposed coastlines for their high-density breeding colonies. The northern fur seal (*Callorhinus ursinus*) currently ranges throughout the North Pacific, with five well-established rookeries on islands spanning the Okhotsk and Bering Seas (Figure 6). Retreating ice is expanding the Arctic passage between the North Pacific and the North Atlantic during the summer months, allowing access to new foraging grounds for many non-ice-dependent species (Figure 6; Kovacs et al., 2011). Such range expansion can result in contact among previously separated populations of marine and terrestrial *Brucella*-carrying species,

contact with existing soil reservoirs of *Brucella*, establishment of new soil reservoirs of *Brucella*, and possible future transmission to other formerly isolated populations. As fur seal foraging grounds expand, there is also potential for the establishment of new rookeries exploiting this new territory. The breeding colony at Bogoslof Island, a volcanic island formed in the Bering Sea during the nineteenth century, was established circa 1980 and has since grown rapidly (Towell and Ream, 2015).

Seals that haul out on ice and land in different parts of their ranges or during different seasons (e.g., gray seals, spotted seals; Figure 6) exhibit reproductive behaviors and population densities that are somewhere between those of the strictly pagophilic and the land-breeding species. This may expose these species to

a combination of the risks outlined above for both ice- and land-pupping pinnipeds.

Variations in ocean temperature and sea ice coverage associated with climate change will alter distributions of marine organisms, both expanding and contracting ranges. These changes in biogeography will alter pathogen transmission within and among populations and species.

“Changes in temperature, pH, O₂, sea ice extent, and many other environmental variables contribute to the impact of climate change on disease in marine systems.”

DISCUSSION

As climate change continues to alter the natural world, species must adapt to survive or be faced with extinction. Average surface temperature is projected to increase by 2°C to 5.4°C within the next century (IPCC, 2007), and our analyses show increasing geographic extent and annual durations of above-critical temperatures. While there exists great variability in host-pathogen interactions, we chose case studies to illustrate some potential effects of increased heat-event durations on host-pathogen dynamics. In particular, we explored:

- **Increased Host Stress.** From our analyses, we predict that extended durations of above-critical temperatures will increase host stress levels and susceptibility to pathogens, potentially leading to local extinctions. We speculate that many marine organisms will become more susceptible to infection with global climate change, as illustrated by the case study of bald sea urchin disease.
- **Increased Pathogen Virulence.** While establishing direct causation between climate change and pathogen virulence may be difficult, cumulative climate

change impacts are likely to increase the virulence of many marine pathogens by influencing factors such as metabolism, growth rate, and migration. Furthermore, sustained elevated temperatures could extend the length of the transmission season, ultimately leading to an increase in the frequency, duration, geographic extent, and intensity of

infections worldwide. The temperature-dependent dynamic of *V. coralliilyticus* infection of *P. damicornis* provides an example of these potential changes.

- **Pathogen Range Expansion.** Range expansions are predicted to become more frequent with increasing sea surface temperatures (Figures 1 and 2). These changes will be particularly evident in the subtropical and temperate bands between 23.5° and 40° latitude, where significant increases in the number of warm months each year will give temperature-constrained pathogens considerable freedom to expand their ranges poleward (Figure 5). This will put unaffected populations at risk, as we expect to see with *P. marinus* infecting previously dermo-free populations of *C. virginica*.
- **Host Range Changes.** Host habitat constriction is likely to increase population densities and thus accelerate rates of intraspecies infection. Habitat expansion, on the other hand, is likely to cause novel contact among populations and disease vectors or reservoirs, potentially increasing interspecies infection. Both of these outcomes

have the capacity to increase the incidence and geographic extent of infection by pathogens in changing physical environments, and can be explored through *Brucella* infection of various Arctic pinnipeds.

Our case studies illustrate how increasing temperatures will have global impacts on host-pathogen relationships, with regionally differentiated outcomes. By 2100, we expect increases of four to eight months of unusually high temperatures in temperate regions, largely due to the predicted migration of temperate zones poleward (Figure 2). Organisms with restricted dispersal (e.g., urchins and oysters) will have difficulty acclimating to temperature changes (Figures 3 and 5). In polar regions, sea ice melt may drastically constrict suitable habitats (Figure 6), while in the tropics, average monthly temperatures (>30°C) may become too high for even warm-water corals (Figure 4). Furthermore, pathogens currently restricted to narrow temperature bands may expand to create greater geographic overlap with host habitats (Figure 5).

Though we have focused on temperature as the dominant climate change forcing due to its ubiquitous impact on coastal and Arctic marine environments and the availability of literature on the subject, temperature will not act in isolation on marine communities: anthropogenic climate change is also causing our ocean to acidify, stratify, and exhibit increased hypoxia. In coastal oceans, there are shifts in freshwater input and sedimentation as a result of changing land-use and precipitation patterns. Moreover, humans are significantly impacting biodiversity through industrial fishing and aquaculture practices (IPCC, 2014).

Not all changes are predicted to have negative consequences for the hosts, and some outcomes may be mixed (Danovaro et al., 2011). Pathogens may be less tolerant of temperature changes than their hosts, resulting in decreased pathogen abundance and local incidence of


infection (Holt et al., 1989; Harvell et al., 2002). Changes in host abundance and distribution may also decrease infection rates by disrupting complex pathogen life cycles, which can rely on multiple hosts, or by reducing host densities below thresholds necessary to support pathogen populations (Grenfell and Dobson, 1995; Auld and Tinsley, 2015).

Changes to host-pathogen relationships not only will affect specific host-pathogen pairs but also will lead to broader impacts in the surrounding ecosystem. For instance, marine viruses are vital to nutrient cycling and are determiners of planktonic species abundance and distribution (Fuhrman, 1999). If current host-pathogen relationships become distorted or new host-pathogen relationships emerge, community composition and subsequent ecosystem services could be greatly altered. Mortalities of key species may also cause ecological phase shifts due to the loss of three-dimensional structures such as corals or kelp. Ecosystem biodiversity and even species evolution can be influenced by marine pathogens (Suttle, 2007). The effects of climate change on marine host-pathogen dynamics can therefore impact every trophic level of marine ecosystems.

Human health may also be affected by these changing dynamics. Pathogen range expansion and increasing virulence may negatively affect economically important species by impacting fisheries productivity and increasing the risk of human foodborne illness. *V. coralliilyticus* strains, for example, have shown resistance to commercially available antibiotics (Vizcaino et al., 2010), and range expansion and horizontal gene transfer may allow this trait to become prevalent in other *Vibrio* species that currently pose no risk to humans. Further investigations of the impact of climate change on *Brucella* and other marine mammal-borne pathogens are necessary, given the potential for transmission both among species of marine mammals and also to human populations.

Changes in host-pathogen relation-

ships may have negative economic impacts on important commercial fisheries. There will be increased stress on many species as pathogen virulence and host mortality accelerate (Karvonen, 2010). Declining abundance or fitness of target species will result in a decline of subsistence fisheries, and developing nations—particularly those in the tropics—are likely to be among the first impacted (Figure 2).

These examples of changes in host-pathogen relationships show the potential for substantial reductions in biodiversity, loss of valuable ecosystem services, and threats to human health. However, there is a great deal of uncertainty associated with making predictions about the impacts of climate change, and marine host-pathogen systems in particular are understudied and little understood in this regard. In order to meaningfully predict, plan for, and mitigate the potential negative consequences of climate change on these host-pathogen systems, it is necessary that we consider the underlying drivers of change and the likely responses of both pathogens and hosts. Complex interactions among multiple factors, such as rising sea surface temperature, anoxia, ocean acidification, and stratification, require further investigation. 

REFERENCES

- Agatsuma, Y. 2007. Ecology of *Strongylocentrotus intermedius*. Pp. 427–441 in *Edible Sea Urchins: Biology and Ecology*, 2nd ed. J.M. Lawrence, ed., Elsevier Science, Amsterdam.
- Auld, S., and M.C. Tinsley. 2015. The evolutionary ecology of complex lifecycle parasites: Linking phenomena with mechanisms. *Heredity* 114(2):125–132, <https://doi.org/10.1038/hdy.2014.84>.
- Banks, P., M. Berrigan, A. Choudhury, L. Craig, D. Diaz, F. Kern, J. King, M. Marshall, L. Robinson, F. Steimle, and others. 2007. *Status Review of the Eastern Oyster (Crassostrea virginica)*. Report to the National Marine Fisheries Service, Northeast Regional Office, Gloucester, MA, USA, NOAA Technical Memo NMFS F/SPO-88, 105 pp.
- Becker, P.T., E. Egea, and I. Eeckhaut. 2008. Characterization of the bacterial communities associated with the bald sea urchin disease of the echinoid *Paracentrotus lividus*. *Journal of Invertebrate Pathology* 98(2):136–147, <https://doi.org/10.1016/j.jip.2007.12.002>.
- Ben-Haim, Y., F.L. Thompson, C.C. Thompson, M.C. Cnockaert, B. Hoste, J. Swings, and E. Rosenberg. 2003a. *Vibrio coralliilyticus* sp. nov., a temperature-dependent pathogen of the coral *Pocillopora damicornis*. *International*

- Journal of Systematic and Evolutionary Microbiology* 53:309–315, <https://doi.org/10.1099/ijs.0.02402-0>.
- Ben-Haim, Y., M. Zicherman-Keren, and E. Rosenberg. 2003b. Temperature-regulated bleaching and lysis of the coral *Pocillopora damicornis* by the novel pathogen *Vibrio coralliilyticus*. *Applied and Environmental Microbiology* 69(7):4,236–4,242, <https://doi.org/10.1128/AEM.69.7.4236-4242.2003>.
- Berkelmans, R. 2002. Time-integrated thermal bleaching thresholds of reefs and their variation on the Great Barrier Reef. *Marine Ecology Progress Series* 229:73–82, <https://doi.org/10.3354/meps229073>.
- Bidegain, G., E.N. Powell, T. Klinck, T. Ben-Horin, and E.E. Hofmann. 2016. Marine infectious disease dynamics and outbreak thresholds: Contact transmission, pandemic infection, and the potential role of filter feeders. *Ecosphere* 7(4):e01286, <https://doi.org/10.1002/ecs2.1286>.
- Boudouresque, C.F., and M. Verlaque. 2007. Ecology of *Paracentrotus lividus*. Pp. 243–285 in *Edible Sea Urchins: Biology and Ecology*, 2nd ed. J.M. Lawrence, ed., Elsevier Science, Amsterdam.
- Bourne, D.G., M. Garren, T.M. Work, E. Rosenberg, G.W. Smith, and C.D. Harvell. 2009. Microbial disease and the coral holobiont. *Trends in Microbiology* 17(12):554–562, <https://doi.org/10.1016/j.tim.2009.09.004>.
- Bourne, D.G., and C.B. Munn. 2005. Diversity of bacteria associated with the coral *Pocillopora damicornis* from the Great Barrier Reef. *Environmental Microbiology* 7(8):1,162–1,174.
- Bower, S.M. 2013. Synopsis of Infectious Diseases and Parasites of Commercially Exploited Shellfish: *Perkinsus marinus* ("Dermo" Disease) of Oysters. Fisheries and Oceans Canada, <http://www.dfo-mpo.gc.ca/science/aah-saa/diseases-maladies/pmdoy-eng.html>.
- Bower, S.M., S.E. McGladdery, and I.M. Price. 1994. Synopsis of infectious diseases and parasites of commercially exploited shellfish. *Annual Review of Fish Diseases* 4(0):1–199, [https://doi.org/10.1016/0959-8030\(94\)90028-0](https://doi.org/10.1016/0959-8030(94)90028-0).
- Brothers, C.J., J. Harianto, J.B. McClintock, and M. Byrne. 2016. Sea urchins in a high-CO₂ world: The influence of acclimation on the immune response to ocean warming and acidification. *Proceedings of the Royal Society B*, <https://doi.org/10.1098/rspb.2016.1501>.
- Burge, C.A., C.M. Eakin, C.S. Friedman, B. Froelich, P.K. Hershberger, E.E. Hofmann, L.E. Petes, K.C. Prager, E. Weil, B.L. Willis, and others. 2014. Climate change influences on marine infectious diseases: Implications for management and society. *Annual Review of Marine Science* 6:249–277, <https://doi.org/10.1146/annurev-marine-010213-135029>.
- Combosch, D.J., and S.V. Vollmer. 2011. Population genetics of an ecosystem-defining reef coral *Pocillopora damicornis* in the tropical eastern Pacific. *PLoS ONE* 6(8): e21200, <https://doi.org/10.1371/journal.pone.0021200>.
- Cook, T., M. Folli, J. Klinck, S. Ford, and J. Miller. 1998. The relationship between increasing sea-surface temperature and the northward spread of *Perkinsus marinus* (Dermo) disease epizootics in oysters. *Estuarine Coastal and Shelf Science* 46(4):587–597, <https://doi.org/10.1006/ecss.1997.0283>.
- Danovaro, R., C. Corinaldesi, A. Dell'Anno, J.A. Fuhrman, J.J. Middelburg, R.T. Noble, and C.A. Suttle. 2011. Marine viruses and global climate change. *FEMS Microbiology Reviews* 35(6):993–1,034, <https://doi.org/10.1111/j.1574-6976.2010.00258.x>.

- D'Croz, L., and J.L. Maté. 2004. Experimental responses to elevated water temperature in genotypes of the reef coral *Pocillopora damicornis* from upwelling and non-upwelling environments in Panama. *Coral Reefs* 23:473–483, <https://doi.org/10.1007/s00338-004-0397-7>.
- de Figueiredo, P., T.A. Ficht, A. Rice-Ficht, C.A. Rossetti, and L.G. Adams. 2015. Pathogenesis and immunobiology of Brucellosis: Review of *Brucella*-host interactions. *The American Journal of Pathology* 185(6):1,505–1,517, <https://doi.org/10.1016/j.ajpath.2015.03.003>.
- de Montaudouin, X., I. Paul-Pont, C. Lambert, P. Gonzalez, N. Raymond, F. Jude, A. Legeay, M. Baudrimont, C. Dang, F. Le Grand, and others. 2010. Bivalve population health: Multistress to identify hot spots. *Marine Pollution Bulletin* 60(8):1,307–1,318, <https://doi.org/10.1016/j.marpolbul.2010.03.011>.
- Doney, S.C., M. Ruckelshaus, J.E. Duffy, J.P. Barry, F. Chan, C.A. English, H.M. Galindo, J.M. Grebmeier, A.B. Hollowed, N. Knowlton, and others. 2012. Climate change impacts on marine ecosystems. *Annual Review of Marine Science* 4:11–37, <https://doi.org/10.1146/annurev-marine-041911-116111>.
- Dove, S., and O. Hoegh-Guldberg. 2006. The cell physiology of coral bleaching. Pp. 55–71 in *Coral Reefs and Climate Change: Science and Management*. J.T. Phinney, O. Hoegh-Guldberg, J. Kleypas, W. Skirving, and A. Strong, eds, American Geophysical Union, Washington, DC.
- Farmanfarmaian, A., and A.C. Giese. 1963. Thermal tolerance and acclimation in the western purple sea urchin, *Strongylocentrotus purpuratus*. *Physical and Biochemical Zoology* 36(3):237–243, <https://doi.org/10.1086/physzool.36.3.30152309>.
- Ford, S.E., and E. Smolowitz. 2007. Infection dynamics of an oyster parasite in its newly expanded range. *Marine Biology* 151(1):119–133, <https://doi.org/10.1007/s00227-006-0454-6>.
- Fuhrman, J.A. 1999. Marine viruses and their biogeochemical and ecological effects. *Nature* 399:541–548, <https://doi.org/10.1038/21119>.
- Garren, M., K. Son, J.B. Raina, R. Rusconi, F. Menolascina, O.H. Shapiro, J. Tout, D.G. Bourne, J.R. Seymour, and R. Stocker. 2014. A bacterial pathogen uses dimethylsulfoniopropionate as a cue to target heat-stressed corals. *ISME Journal* 8(5):999–1,007, <https://doi.org/10.1038/ismej.2013.210>.
- Garren, M., K. Son, J. Tout, J.R. Seymour, and R. Stocker. 2016. Temperature-induced behavioral switches in a bacterial coral pathogen. *ISME Journal* 10(6):1,363–1,372, <https://doi.org/10.1038/ismej.2015.216>.
- Girard, D., S. Clemente, K. Toledo-Guedes, A. Brito, and J.C. Hernandez. 2012. A mass mortality of subtropical intertidal populations of the sea urchin *Paracentrotus lividus*: Analysis of potential links with environmental conditions. *Marine Ecology* 33(3):377–385, <https://doi.org/10.1111/j.1439-0485.2011.00491.x>.
- Gnanadesikan, A., K.W. Dixon, S.M. Griffies, V. Balaji, M. Barreiro, J.A. Beesley, W.F. Cooke, T.L. Delworth, R. Gerdes, M.J. Harrison, and others. 2006. GFDL's CM2 global coupled climate models: Part II. The baseline ocean simulation. *Journal of Climate* 19(5):675–697, <https://doi.org/10.1175/JCLI3630.1>.
- Grenfell, B.T., and A.P. Dobson, eds. 1995. *Ecology of Infectious Diseases in Natural Populations*. Cambridge University Press, 536 pp.
- Harvell, C.D., C.E. Mitchell, J.R. Ward, S. Altizer, A.P. Dobson, R.S. Ostfeld, and M.D. Samuel. 2002. Climate warming and disease risks for terrestrial and marine biota. *Science* 296(5576):2,158–2,162, <https://doi.org/10.1126/science.1063699>.
- Heron, S.F., J.A. Maynard, R. van Hooidin, and C.M. Eakin. 2016. Warming trends and bleaching stress of the world's coral reefs 1985–2012. *Scientific Reports* 6:38402, <https://doi.org/10.1038/srep38402>.
- Hoeksema, B.W., A. Rogers, and M.C. Quibilan. 2014. *Pocillopora damicornis*. The IUCN Red List of Threatened Species, <https://doi.org/10.2305/IUCN.UK.2014-1.RLTS.T133222A54216898.en>.
- Hughes, T.P., J.T. Kerry, M. Alvarez-Noriega, J.G. Alvarez-Romero, K.D. Anderson, A.H. Baird, R.C. Babcock, M. Beger, D.R. Bellwood, R. Berkelmans, and others. 2017. Global warming and recurrent mass bleaching of corals. *Nature* 543(7645):373–377, <https://doi.org/10.1038/nature21707>.
- Holt, R.A., A. Amandi, J.S. Rohovec, and J.L. Fryer. 1989. Relation of water temperature to bacterial cold-water disease in coho salmon, Chinook salmon, and rainbow trout. *Journal of Aquatic Animal Health* 1(2):94–101, [https://doi.org/10.1577/1548-8667\(1989\)001<0094:ROWTTB>2.3.CO;2](https://doi.org/10.1577/1548-8667(1989)001<0094:ROWTTB>2.3.CO;2).
- IPCC. 2007. *Climate Change 2007: Synthesis Report. Contribution of Working Groups I, II and III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Core Writing Team, R.K. Pachauri, and A. Reisinger, eds, IPCC, Geneva, Switzerland, 104 pp.
- IPCC. 2014. *Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Core Writing Team, R.K. Pachauri, and L.A. Meyer, eds, IPCC, Geneva, Switzerland, 151 pp.
- Jangoux, M. 1987. Diseases of Echinodermata: Part 1. Agents, microorganisms and protists. *Diseases of Aquatic Organisms* 2(2):147–162, <https://doi.org/10.3354/dao002147>.
- Jangoux, M. 1990. Diseases of Echinodermata. Chapter 5 in *Diseases of Marine Animals. Volume III. Introduction. Cephalopoda, Annelida, Crustacea, Chaetognatha, Echinodermata, Urochordata*. O. Kinne, ed., Biologische Anstalt Helgoland, Hamburg.
- Jenkins, C., P. Hick, M. Gabor, Z. Spiers, S.A. Fell, X.N. Gu, A. Read, J. Go, M. Dove, W. O'Connor, and others. 2013. Identification and characterization of an ostreid herpesvirus-1 microvariant (OsHV-1 mu-var) in *Crassostrea gigas* (Pacific oysters) in Australia. *Diseases of Aquatic Organisms* 105(2):109–126, <https://doi.org/10.3354/dao02623>.
- Karvonen, A., P. Rintamaki, J. Jokela, and E.T. Valtonen. 2010. Increasing water temperature and disease risks in aquatic systems: Climate change increases the risk of some, but not all, diseases. *International Journal for Parasitology* 40(13):1,483–1,488, <https://doi.org/10.1016/j.ijpara.2010.04.015>.
- Kimes, N.E., C.J. Grim, W.R. Johnson, N.A. Hasan, B.D. Tall, M.H. Kothary, H. Kiss, A.C. Munk, R. Tapia, R. Green, and others. 2012. Temperature regulation of virulence factors in the pathogen *Vibrio coralliilyticus*. *ISME Journal* 6(4):835–846, <https://doi.org/10.1038/ismej.2011.154>.
- Kordas, R.L., C.D.G. Harley, and M.I. O'Connor. 2011. Community ecology in a warming world: The influence of temperature on interspecific interactions in marine systems. *Journal of Experimental Marine Biology and Ecology* 400(1–2):218–226, <https://doi.org/10.1016/j.jembe.2011.02.029>.
- Kovacs, K.M., C. Lydersen, J.E. Overland, and S.E. Moore. 2011. Impacts of changing sea-ice conditions on Arctic marine mammals. *Marine Biodiversity* 41(1):181–194, <https://doi.org/10.1007/s12526-010-0061-0>.
- La Peyre, M.K., S.M. Casas, A. Villalba, and J.F. La Peyre. 2008. Determination of the effects of temperature on viability, metabolic activity and proliferation of two Perkinsus species, and its significance to understanding seasonal cycles of perkinsosis. *Parasitology* 135(4):505–519, <https://doi.org/10.1017/S0031182008004150>.
- Lambourn, D., M. Garner, D. Ewalt, S. Raverty, I. Sidor, S.J. Jeffries, J. Rhyhan, and J.K. Gaydos. 2013. *Brucella pinnipedialis* infections in Pacific harbor seals (*Phoca vitulina Richardsi*) from Washington state, USA. *Journal of Wildlife Diseases* 49(4):802–815, <https://doi.org/10.7589/2012-05-137>.
- Lester, S.E., E.D. Tobin, and M.D. Behrens. 2007. Disease dynamics and the potential role of thermal stress in the sea urchin, *Strongylocentrotus purpuratus*. *Canadian Journal of Fisheries and Aquatic Sciences* 64(2):314–323, <https://doi.org/10.1139/f07-010>.
- Maes, P., and M. Jangoux. 1984. The bald-sea-urchin disease: A biopathological approach. *Helgolander Meeresuntersuchungen* 37(1–4):217–224, <https://doi.org/10.1007/BF01989306>.
- Maynard, J., R. van Hooidin, C.D. Harvell, C.M. Eakin, G. Liu, B.L. Willis, G.J. Williams, M.L. Groner, A. Dobson, S.F. Heron, and others. 2016. Improving marine disease surveillance through sea temperature monitoring, outlooks and projections. *Philosophical Transactions of the Royal Society B* 371:20150208, <https://doi.org/10.1098/rstb.2015.0208>.
- Meehl, G.A., C. Covey, K.E. Taylor, T. Delworth, R.J. Stouffer, M. Latif, B. McAvaney, and J.F. Mitchell. 2007. The WCRP CMIP3 multi-model dataset: A new era in climate change research. *Bulletin of the American Meteorological Society* 88(9):1,383–1,394, <https://doi.org/10.1175/BAMS-88-9-1383>.
- Nymo, I.H., M. Tryland, and J. Godfroid. 2011. A review of *Brucella* infection in marine mammals, with special emphasis on *Brucella pinnipedialis* in the hooded seal (*Cystophora cristata*). *Veterinary Research* 42:13, <https://doi.org/10.1186/1297-9716-42-93>.
- Pollock, F.J., B. Wilson, W.R. Johnson, P.J. Morris, B.L. Willis, and D.G. Bourne. 2010. Phylogeny of the coral pathogen *Vibrio coralliilyticus*. *Environmental Microbiology Reports* 2(1):172–178, <https://doi.org/10.1111/j.1758-2229.2009.00131.x>.
- Pompa, S., P.R. Ehrlich, and G. Ceballos. 2011. Global distribution and conservation of marine mammals. *Proceedings of the National Academy of Sciences of the United States of America* 108(38):13,600–13,605, <https://doi.org/10.1073/pnas.1101525108>.
- Roekner, E., R. Brokopf, M. Esch, M. Giorgetta, S. Hagemann, L. Kornblueh, E. Manzini, U. Schlese, and U. Schulzweida. 2004. *The Atmospheric General Circulation Model ECHAM5 Part II*. Max-Planck Institut für Meteorologie, Hamburg, Germany, 127 pp.
- Scheibling, R.E., and R.L. Stephenson. 1984. Mass mortality of *Strongylocentrotus droebachiensis* (echinodermata, echinoidea) off Nova Scotia, Canada. *Marine Biology* 78(2):153–164, <https://doi.org/10.1007/BF00394695>.
- Scholz, H.C., Z. Hubalek, J. Nesvadbova, H. Tomaso, G. Vergnaud, P. Le Fleche, A.M. Whatmore, S. Al Dahouk, M. Kruger, C. Lodri, and M. Pfeffer. 2008. Isolation of *Brucella microti* from soil. *Emerging Infectious Diseases* 14(8):1,316–1,317, <https://doi.org/10.3201/eid1408.080286>.
- Segarra, A., J.F. Pepin, I. Arzul, B. Morga, N. Faury, and T. Renault. 2010. Detection and description of a particular Ostreid herpesvirus 1 genotype associated with massive mortality outbreaks of Pacific oysters, *Crassostrea gigas*, in France in 2008. *Virus Research* 153(1):92–99, <https://doi.org/10.1016/j.virusres.2010.07.011>.

- Simpkins, M.A., L.M. Hiruki-Raring, G. Sheffield, J.M. Grebmeier, and J.L. Bengtson. 2003. Habitat selection by ice-associated pinnipeds near St. Lawrence Island, Alaska in March 2001. *Polar Biology* 26(9):577–586, <https://doi.org/10.1007/s00300-003-0527-7>.
- Soudant, P., F.L.E. Chu, and A. Volety. 2013. Host-parasite interactions: Marine bivalve molluscs and protozoan parasites, *Perkinsus* species. *Journal of Invertebrate Pathology* 114(2):196–216, <https://doi.org/10.1016/j.jip.2013.06.001>.
- Spickler, A.R. 2009. *Brucellosis in Marine Mammals*. Last updated July 2009, 7 pp., http://www.cfsph.iastate.edu/Factsheets/pdfs/brucellosis_marine.pdf.
- Sunila, I. 2007. Dermo Disease, <http://www.ct.gov/doag/lib/doag/aquaculture/dermo.pdf>.
- Suttle, C.A. 2007. Marine viruses: Major players in the global ecosystem. *Nature Reviews Microbiology* 5(10):801–812, <https://doi.org/10.1038/nrmicro1750>.
- Sweet, M., M. Bulling, and J.E. Williamson. 2016. New disease outbreak affects two dominant sea urchin species associated with Australian temperate reefs. *Marine Ecology Progress Series* 551:171–183, <https://doi.org/10.3354/meps11750>.
- Thakur, S.D., R.K. Vaid, A.K. Panda, and Y. Saini. 2012. Marine mammal brucellosis: A new dimension to an old zoonosis. *Current Science* 103(8):902–910.
- The Center for Food Security & Public Health. 2009. *Brucellosis in Marine Mammals*. The Center for Food Security & Public Health, Iowa State University, 7 pp., http://www.cfsph.iastate.edu/Factsheets/pdfs/brucellosis_marine.pdf.
- Thompson, C.C., F.L. Thompson, A.C.P. Vicente, and J. Swings. 2007. Phylogenetic analysis of *Vibrios* and related species by means of *atpA* gene sequences. *International Journal of Systematic and Evolutionary Microbiology* 57:2,480–2,484, <https://doi.org/10.1099/ijs.0.65223-0>.
- Tout, J., N. Siboni, L.F. Messer, M. Garren, R. Stocker, N.S. Webster, P.J. Ralph, and J.R. Seymour. 2015. Increased seawater temperature increases the abundance and alters the structure of natural *Vibrio* populations associated with the coral *Pocillopora damicornis*. *Frontiers in Microbiology* 6:432, <https://doi.org/10.3389/fmicb.2015.00432>.
- Towell, R., and R. Ream. 2015. *Bogoslof Island Research: 2015*. National Oceanic and Atmospheric Administration, NOAA Alaska Fisheries Science Center Quarterly Report.
- Vezzulli, L., R.R. Colwell, and C. Pruzzo. 2013. Ocean warming and spread of pathogenic *Vibrios* in the aquatic environment. *Microbial Ecology* 65(4):817–825, <https://doi.org/10.1007/s00248-012-0163-2>.
- Vizcaino, M.I., W.R. Johnson, N.E. Kimes, K. Williams, M. Torralba, K.E. Nelson, G.E. Smith, E. Weil, P.D.R. Moeller, and P.J. Morris. 2010. Antimicrobial resistance of the coral pathogen *Vibrio coralliilyticus* and Caribbean sister phylotypes isolated from a diseased octocoral. *Microbial Ecology* 59(4):646–657, <https://doi.org/10.1007/s00248-010-9644-3>.
- Ward, J.R., and K.D. Lafferty. 2004. The elusive baseline of marine disease: Are diseases in ocean ecosystems increasing? *PLoS Biology* 2(4):542–547, <https://doi.org/10.1371/journal.pbio.0020120>.
- Xu, F., N. Gonzalez-Escalona, J. Haendiges, R.A. Myers, J. Ferguson, T. Stiles, E. Hickey, M. Moore, J.M. Hickey, C. Schillaci, and others. 2017. Sequence type 631 *Vibrio parahaemolyticus*, an emerging foodborne pathogen in North America. *Journal of Clinical Microbiology* 55(2):645–648, <https://doi.org/10.1128/JCM.02162-16>.
- Young, B.G., S.H. Ferguson, and N.J. Lunn. 2015. Variation in ringed seal density and abundance in western Hudson Bay estimated from aerial surveys, 1995 to 2013. *Arctic* 68(3):301–309, <https://doi.org/10.14430/arctic4503>.

ACKNOWLEDGMENTS

The topic of this paper was assigned as a term project to SIO280, the graduate-level introductory biological oceanography course at Scripps Institution of Oceanography, taught by Peter Franks. The 39 students, representing all the disciplines at Scripps, submitted nine group papers that were subsequently synthesized into this paper by the named authors, listed alphabetically. The remaining 28 students were: Corinne Amir, Isabella Arzeno, Katherine Bauman, Kayla Blincow, Katrina Cantu, Pierre Churukian, Kaitlin Creamer, Ana Villas Boas, Jordan DiNardo, Julia Dohner, Elizabeth Fine, Kayleen Fulton, Dulce Guillen Matus, Heena Joo, Sho Kodera, Pichaya Lertvilai, Savannah Lewis, Caroline Lowcher, Vivian Luong, Jeremiah Minich, Kelli Mullane, Anai Novoa, Ariel Rabines, Hugh Runyan, Mohammad Sederat, Derek Smith, Cailan Sugano, and Summer Webb. We thank two anonymous reviewers and Shang-Ping Xie for very helpful comments on earlier versions of the manuscript.

AUTHORS

Rebecca E. Cohen, Chase C. James, Allison Lee, Maurizio M. Martinelli, Wendy T. Muraoka, Mikayla Ortega, Rachel Sadowski, Lark Starkey, Angela R. Szesciorka, Sonya E. Timko, and Elliot L. Weiss were students in biological oceanography class SIO280, and Peter J.S. Franks (pfranks@ucsd.edu) is Professor, Scripps Institution of Oceanography, University of California, San Diego, La Jolla, CA, USA.

ARTICLE CITATION

Cohen, R.E., C.C. James, A. Lee, M.M. Martinelli, W.T. Muraoka, M. Ortega, R. Sadowski, L. Starkey, A.R. Szesciorka, S.E. Timko, E.L. Weiss, and P.J.S. Franks. 2018. Marine host-pathogen dynamics: Influences of global climate change. *Oceanography* 31(2):182–193, <https://doi.org/10.5670/oceanog.2018.201>.